

1 **SUPPLEMENTARY DATA**

	1	2	3	4	5	6	7	8	9	10	11	12
A	EEKA	EEKA	EEKA		BW	BW	BW		JW	JW	JW	
B	EEKA amp	EEKA amp	EEKA amp		BW amp	BW amp	BW amp		JW amp	JW amp	JW amp	
C	EEKA kan	EEKA kan	EEKA kan		BW kan	BW kan	BW kan					
D												
E	EEKA neg	EEKA neg	EEKA neg		BW neg	BW neg	BW neg		JW neg	JW neg	JW neg	
F	EEKA amp neg	EEKA amp neg	EEKA amp neg		BW amp neg	BW amp neg	BW amp neg		JW amp neg	JW amp neg	JW amp neg	
G	EEKA kan neg	EEKA kan neg	EEKA kan neg		BW kan neg	BW kan neg	BW kan neg					
H									Blank	Blank	Blank	

2 **Figure S1. Plate set-up of 96-well absorbance-based lysis assay showing *E. coli* strains**

3 **and type of antibiotic.** Rows E-G contain negative controls (no phage) used to construct

4 growth curves. Row A contains the “no antibiotic” control. EEKA = *E. coli* EEKA18-1, BW

5 = *E. coli* BW25113, JW = *E. coli* JW5437, amp = ampicillin, kan = kanamycin, neg =

6 negative control.

7

A

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1  NNNNNNNNNN NNNNCCGATG GGCATCGGAC CTTTATTGT GCACAGAAAA
51  GGCCAGCCTC GCTTGAGACT GGCCTTTCTG ACAGATGCTT ACTTACTCGC
101 GGAACAGCGC TTCTGTAGGC TGGAGCTGCT TCGAAGTTCC TATACTTTCT
151 AGAGAATAGG AACTTCGAAC TGCAGGTCGA CGGATCCCGA ATCAAATCGT
201 TATCACTGGG TTCTGTCT ACTAAGGCCT TTTCGTCAA AACCTCAACT
251 CGTTCTCAT CAAATCCGC ATCTTCATT AAATCATGAA CTTTCAGCGT
301 ATTCTGACTC ATAAGGTGGC TCCTACCCGT GATCCCTTGA CGGAACATTC
351 AAGCAAAAGC CTGGTCCGC CGATNNNNNC NTGGCGGCAA A

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B

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ALIGNMENTS
>CP009273.1 Escherichia coli BW25113, complete genome
Length=4631469

ALIGNMENT 1:

Score = 42.1 bits (21), Expect = 8e-06
Identities = 21/21 (100%), Gaps = 0/21 (0%)
Strand=Plus/Plus

Query 1          TTACTCGCGGAACAGCGCTTC 21
                |||
Sbjct 2859918    TTACTCGCGGAACAGCGCTTC 2859938

ALIGNMENT 2:

Score = 217 bits (117), Expect = 2e-57
Identities = 117/117 (100%), Gaps = 0/117 (0%)
Strand=Plus/Plus

Query 5          CAAATCGTTATCACTGGGTTCTGTTCTACTAAGGCCTTTTCGTCAAAAACCTCAACTCC 64
                |||
Sbjct 2860791    CAAATCGTTATCACTGGGTTCTGTTCTACTAAGGCCTTTTCGTCAAAAACCTCAACTCC 2860850

Query 65         GTTCTCATCAAATCCGCATCTTCATTTAAATCATGAACTTTCAGCGTATTCTGACT 121
                |||
Sbjct 2860851    GTTCTCATCAAATCCGCATCTTCATTTAAATCATGAACTTTCAGCGTATTCTGACT 2860907

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8

9 **Figure S2. Sanger sequencing of the colony #1 amplicon shows the presence of a partial**

10 **‘scar’ sequence after kanamycin resistance cassette removal.** The sequence of the purified

11 PCR product was mailed to GeneWiz for Sanger sequencing using the *rpoS* forward primer.

12 Note that *rpoS* lies on the negative strand of the *E. coli* BW25113 genome. (A) Raw

13 sequencing results (5’ to 3’). The nucleotide sequence of the stop codon is italicized and

14 bolded. 18 nucleotides that correspond to 6 C-terminal amino acids are italicized. The partial

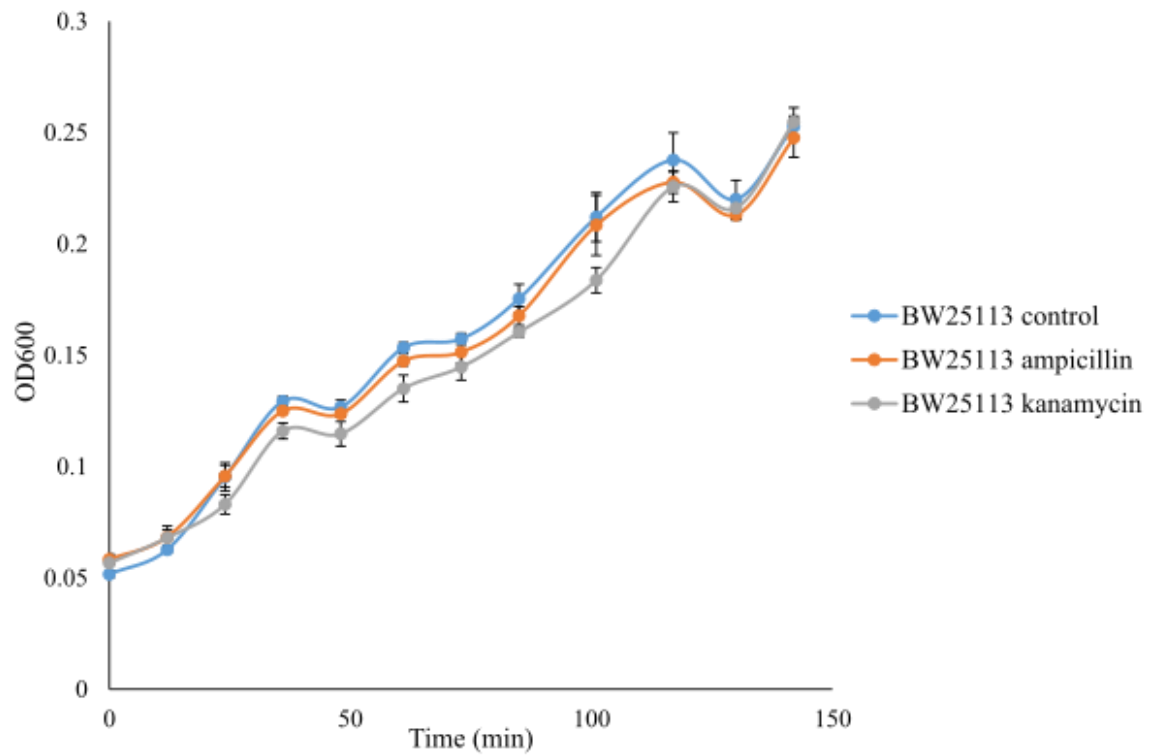
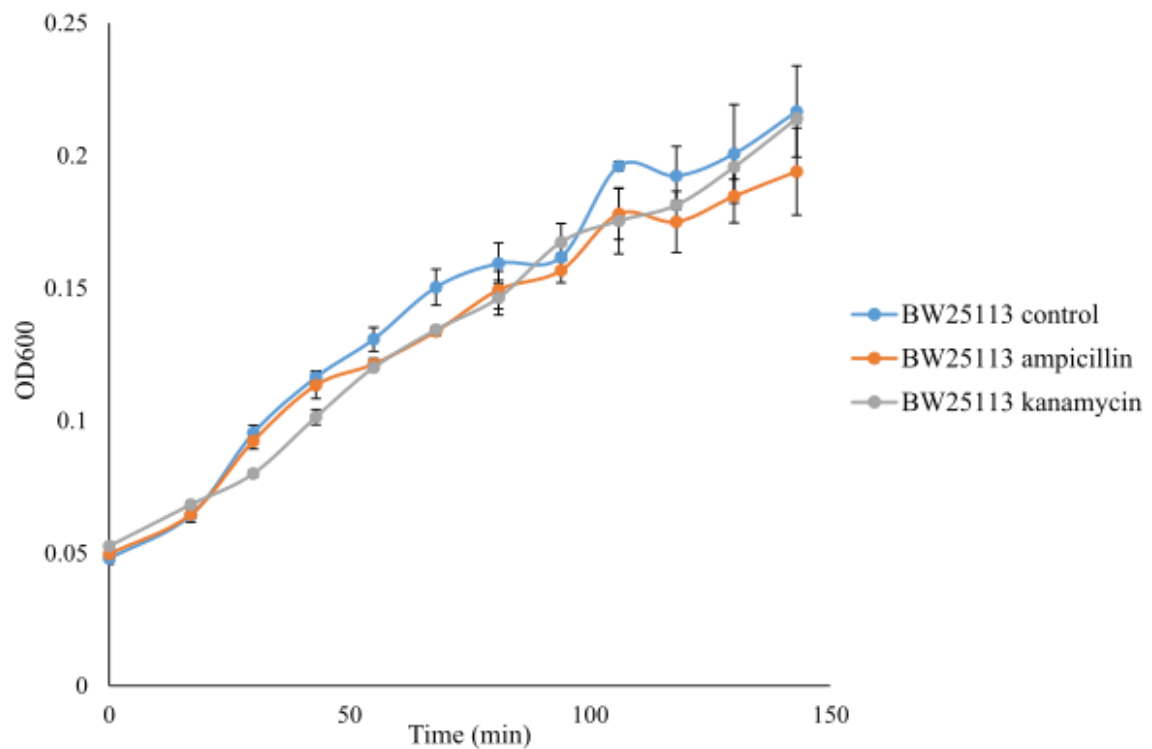
15 ‘scar’ sequence is bolded. The 121 unexpected nucleotides are underlined. The sequence

16 corresponding to the start codon is bolded and underlined. (B) BLASTN 2.8.0+ alignment

17 results. Alignment 1 shows the 18 nucleotides present in the 3’ region of *rpoS* as well as the

18 stop codon nucleotide sequence aligns with *rpoS*. Alignment 2 shows 117 nucleotides out of

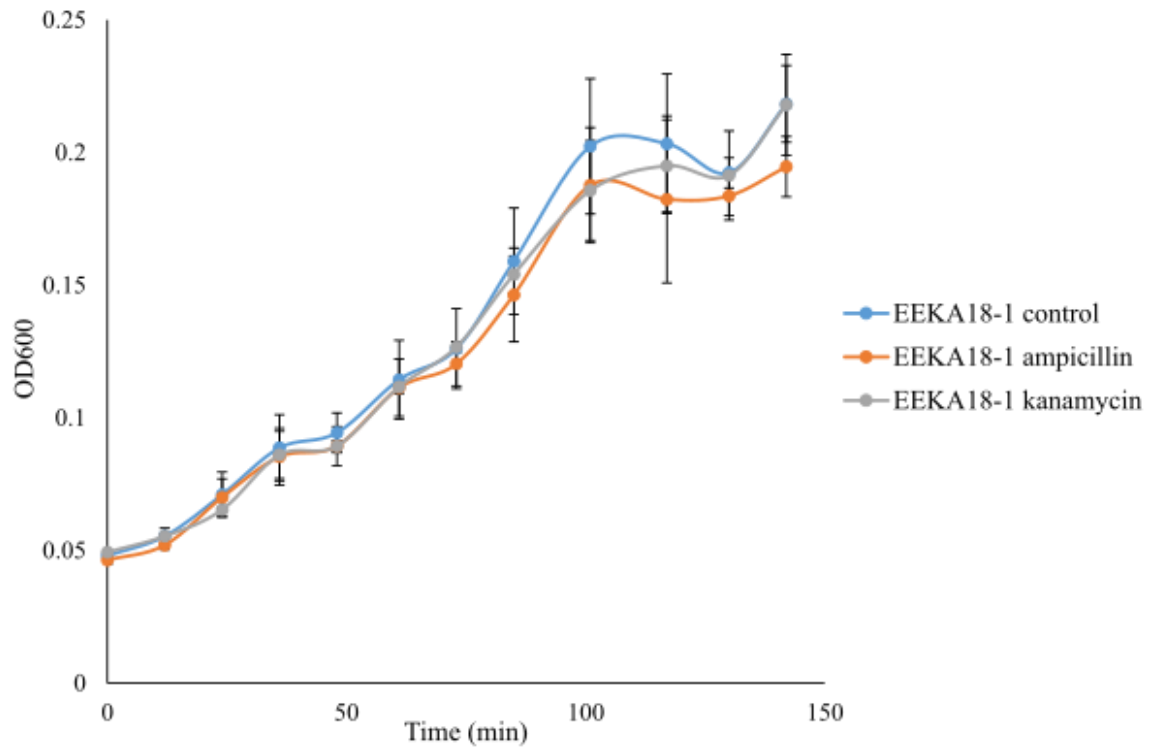
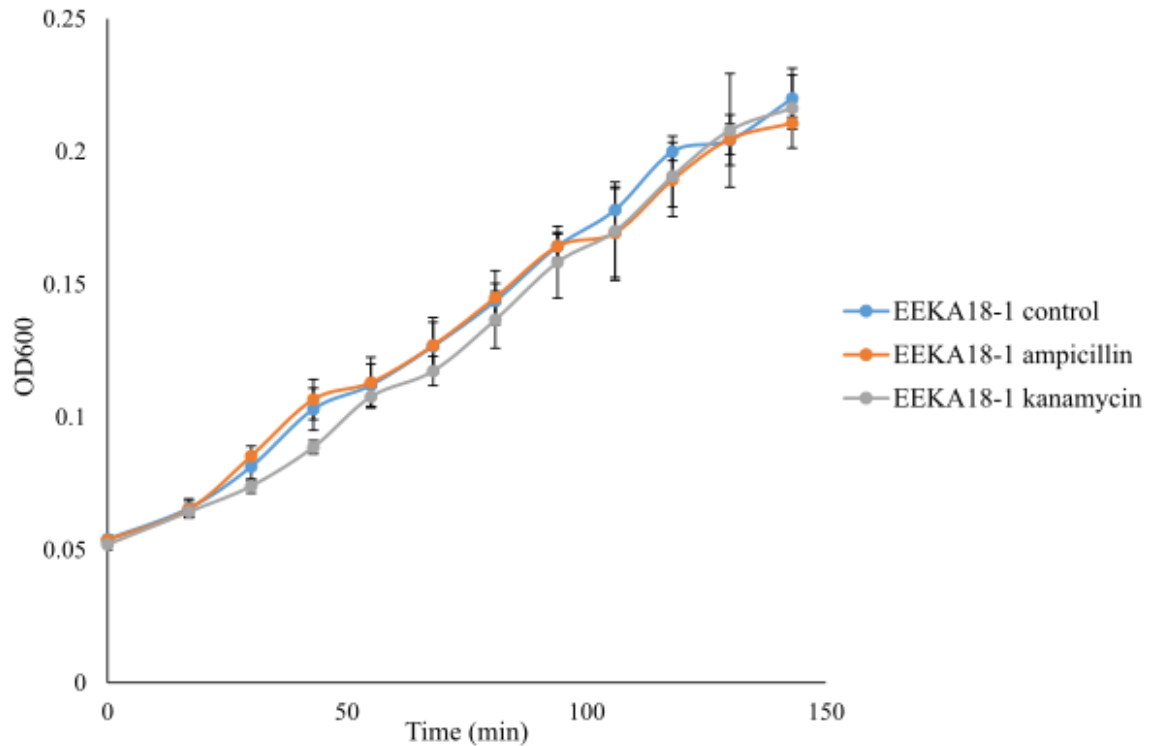
19 the extra 121 nucleotides align with a sequence in the 5’ region of *rpoS*.

A**B**

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21 **Figure S3. The growth rate of *E. coli* BW25113 pre-treated with sub-lethal**22 **concentrations of ampicillin or kanamycin is similar to untreated BW25113.** Individual23 graphs in A and B represent growth curves from two biological replicates. *E. coli* BW25113

24 was cultured overnight in three media conditions: LB control, LB with sub-lethal
25 concentrations of kanamycin, or LB with sub-lethal concentrations of ampicillin. The cells
26 were subcultured to obtain log-phase cells. The cultures were normalized to an OD₆₀₀ of 0.3,
27 plated in triplicates on a 96-well plate, and incubated at 37°C. OD₆₀₀ measurements were
28 taken in 10-minute intervals using a BioTek Epoch Microplate Spectrophotometer to generate
29 the growth curves. Error bars represent standard deviations.

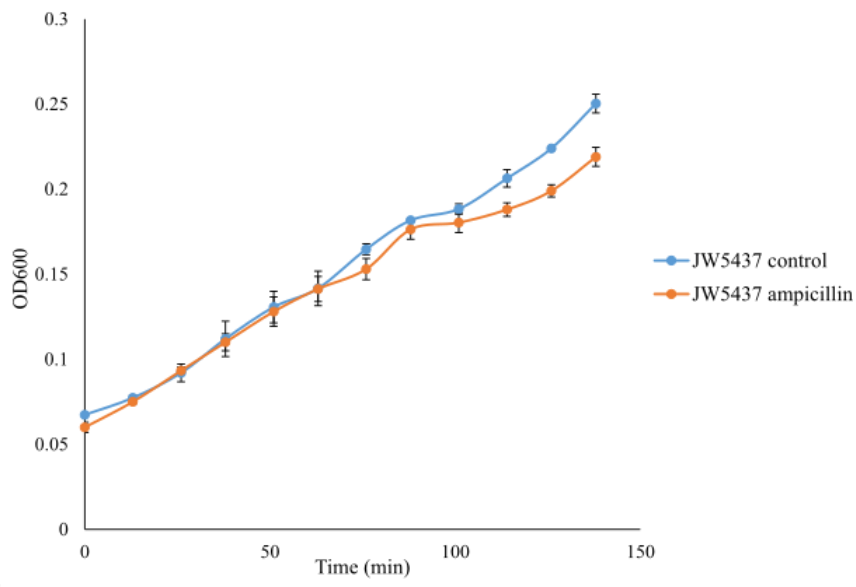
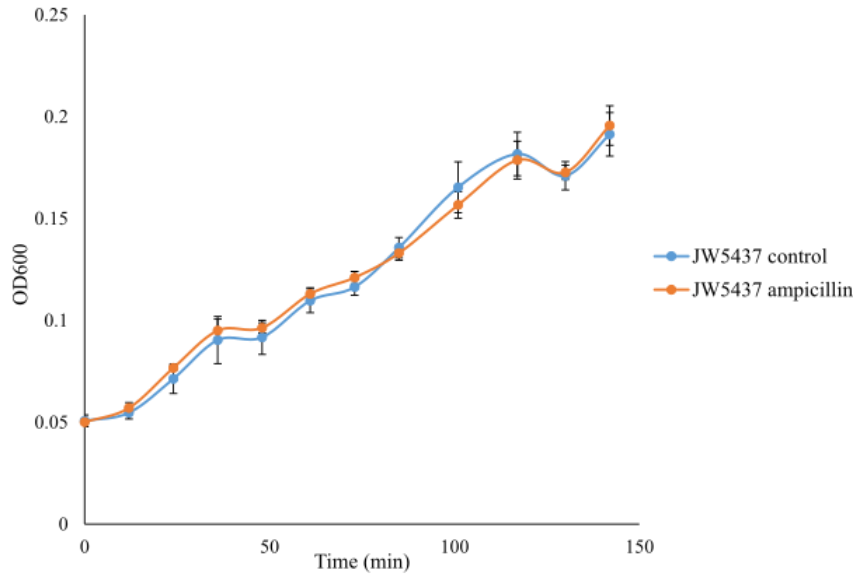
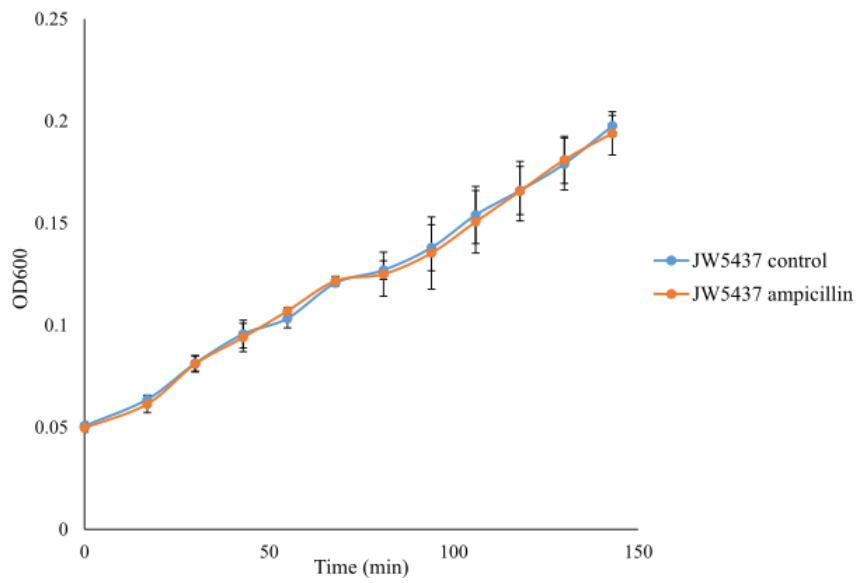
A**B**

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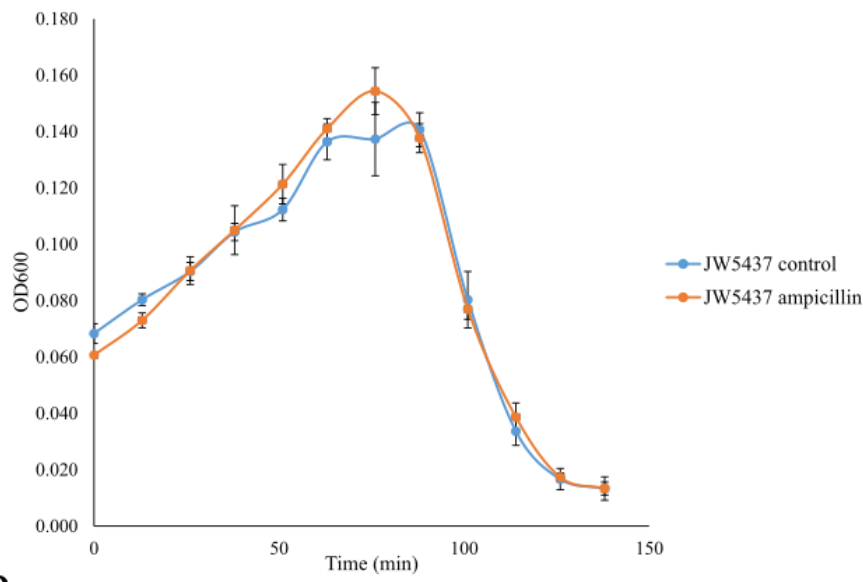
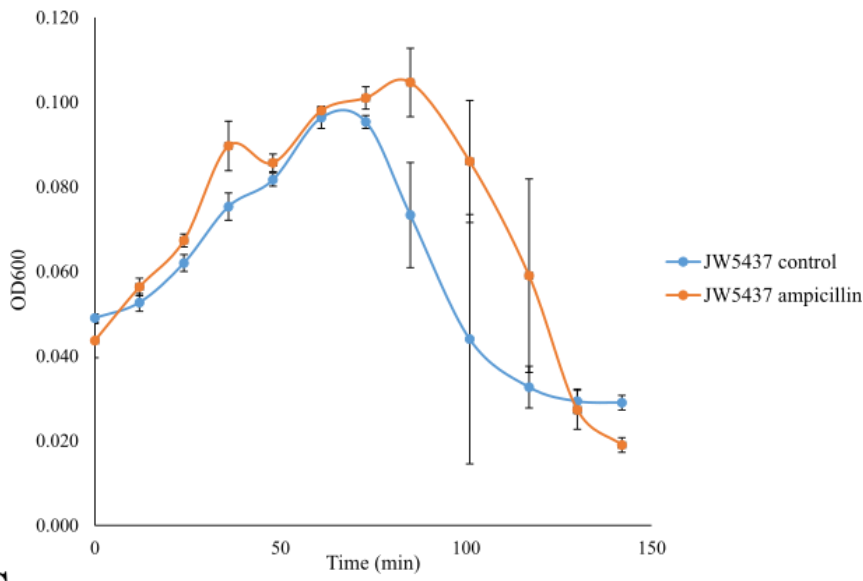
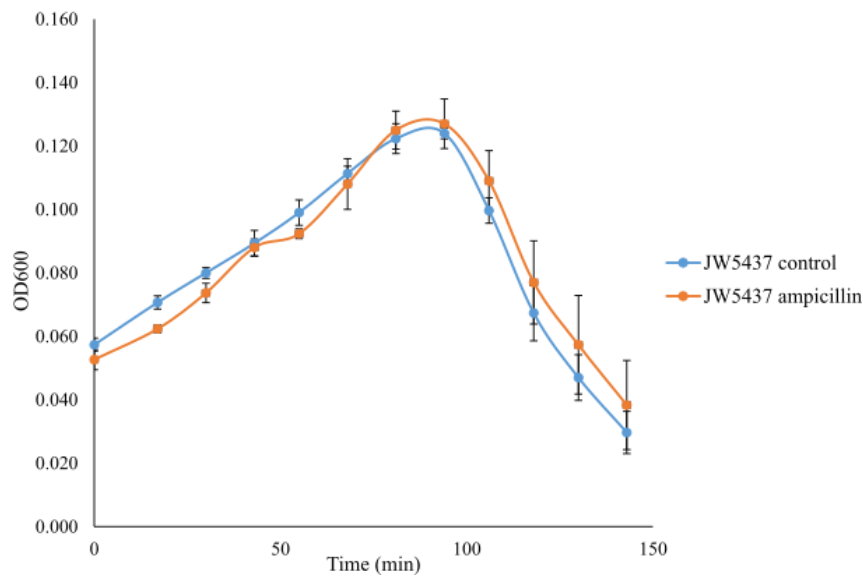
31 **Figure S4. The growth rate of *E. coli* EEKA18-1 pre-treated with sub-lethal**
32 **concentrations of ampicillin or kanamycin is similar to untreated EEKA18-1.** Individual
33 graphs in A and B represent growth curves from two biological replicates. *E. coli* EEKA18-1

34 was cultured overnight in three media conditions: LB control, LB with sub-lethal
35 concentrations of kanamycin, or LB with sub-lethal concentrations of ampicillin. The cells
36 were subcultured to obtain log-phase cells. The cultures were normalized to an OD₆₀₀ of 0.3,
37 plated in triplicates on a 96-well plate, and incubated at 37°C. OD₆₀₀ measurements were
38 taken in 10 minute intervals using a BioTek Epoch Microplate Spectrophotometer to generate
39 the growth curves. Error bars represent standard deviations.

40

A**B****C**

42 **Figure S5. The growth rate of *E. coli* JW5437 pre-treated with sub-lethal concentrations**
43 **of ampicillin is similar to untreated JW5437.** Individual graphs in A, B, and C represent
44 growth curves from three biological replicates. *E. coli* BW25113 was cultured overnight in
45 two media conditions: LB control or LB with sub-lethal concentrations of ampicillin. The
46 cells were subcultured to obtain log-phase cells. The cultures were normalized to an OD₆₀₀ of
47 0.3, plated in triplicates on a 96-well plate, and incubated at 37°C. OD₆₀₀ measurements were
48 taken in 10 minute intervals using a BioTek Epoch Microplate Spectrophotometer to generate
49 the growth curves. Error bars represent standard deviation.

A**B****C**

51 **Figure S6. Pre-treatment of *E. coli* JW5437 with sub-lethal concentrations of ampicillin**
52 **does not delay bacteriophage T7-mediated cell lysis.** Individual graphs in A, B, and C
53 represent lysis curves from three biological replicates. *E. coli* JW5437 was grown overnight
54 in two media conditions: LB broth or LB broth with sub-lethal concentrations of ampicillin.
55 The cells were subcultured the next day and grown to log phase. The cultures were
56 normalized to an OD₆₀₀ of 0.3, and 90 μL of each culture was plated in triplicates in a 96-well
57 plate. A total of 10 μL of diluted T7 bacteriophage was added to the cells to obtain a MOI of
58 0.05. Negative controls were plated in triplicates using 10 μL of LB broth instead of
59 bacteriophage. The plated cells were incubated at 37°C, and the OD₆₀₀ was read in a plate
60 reader in 10 minute intervals until the OD₆₀₀ dipped below the starting values. Error bars
61 represent standard deviations.